

Application of molecular methods in weed science

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Abstract

Molecular methods are useful tools for weed science, especially in the area of weed resistance to herbicides and gene flow from herbicide tolerant crops to their wild relatives. Also, genetic variability plays an important role in weed susceptibility to herbicides and affect on strategies of control. For all of these studies, DNA, as a starting material, could be extracted by various methods; though, the easiest and the most suitable is extraction by using commercially available kits. The most important part of molecular analysis is selection and design of adequate primers for successful DNA amplification. Usually, primer selection and designing are based on DNA sequences stored in GenBank. Analysis following selected DNA fragments will depend on type of research. For weed resistance or gene flow studies, amplified fragments are sequenced and obtained information compared with the GenBank sequence database, with the aim to check for mutation(s) presence. For genetic diversity of weed species analysis of amplified DNA fragments include Capillary Electrophoresis.

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1 Introduction

Molecular methods can be useful for different weed science research topics including molecular determination of weed species which is difficult for determination based on non-molecular methods, population variability, weed resistance to herbicides and gene flow between herbicide-tolerant crops and their wild relatives.

Over the last period weed resistance to herbicides has become an increasing problem (Moss et al. 2007; Michitte et al. 2007). In most cases, evolved weed resistance is due to mutation/mutations within gene encoding enzymes which represent herbicide target site. Therefore, it is possible to use a variety of molecular-based assays that are much faster and less labor intensive than traditional methods (e.g. whole-plant bioassay). Detecting weed resistance to herbicides using DNA based techniques is a very important mission, especially with increasing use of newly-bred herbicide-tolerant crops. There are potential risks associated with growing these crops such as gene flow from herbicide-tolerant crops to non-tolerant crops, or to wild relatives, or volunteer crops. This leads to incidences of resistant species (weeds) (Martinez-Ghersa et al. 1997). Molecular markers have proven valuable in determining the frequency of crop-weed hybridization.

Molecular-based approaches have been used in a variety of ways to explore the genetic diversity of weeds. Studies of genetic diversity can be used for determination of species center of origin (Goolsby et al. 2006, Madeira et al. 2007). Such knowledge can be used to direct searches for potential biological control agents (Paterson et al. 2009). Other genetic diversity studies have been conducted with vegetative propagated perennial weed species, with a goal to determine the relative role of sexual versus vegetative reproduction to the success of the weeds (Slotta et al. 2006). Also, studies of genetic diversity in weed populations can be extremely important because they provide essential background for their different susceptibility to herbicides.

2 Materials, Methods and Notes

2.1 Assessment of gene flow from herbicide tolerant sunflower to weedy sunflower by end-point PCR

End-point PCR is suitable for detection mutations responsible for weed resistance to herbicides and confirmation gene flow from tolerant crops to weedy relatives. Namely, the alteration of ALS (acetolactate synthase) gene by one of many possible point mutations is main mechanism of weed resistance to ALS-inhibiting herbicides, which represent group of herbicides to which weeds usually developed resistance. There are eight possible point mutations detected until now in different weed species. As position of potential mutation is known,

their detection is based on amplification of the appropriate DNA fragment, using specific primers. After amplification PCR products have to be sequenced and check presence of mutation in obtained sequences.

2.1.1 Plant material

Seed material for weed resistance to herbicides research should be collected in the fields for which there are indications about resistance development. Also, it is necessary to collect seeds of the same species from the areas where there is no herbicide application history. For gene flow studies, it is possible to establish field experiment which includes different variants of crop-wild relative distance or to collect seeds from wild relatives of crop (in our case weedy sunflower seeds) from the tolerant crop growing area. Young plants produced from collected seeds are sampled and some leaf samples (taken from a single plant) were used for DNA extraction immediately after sampling, while some of them stored in a freezer (-20°C) until analysis. Before analysis samples were lyophilized after storage at the -80°C during 24h.

2.1.2 DNA extraction

DNA extraction was done using the QiagenDneasy[®] Plant Mini Kit following the manufacturer's protocol (<https://www.qiagen.com/dz/shop/sample-technologies/dna/dna-preparation/dneasy-plant-mini-kit/>). The quality and concentration of extracted DNA were determined spectrophotometrically using a Nanodrop[®] 1000. DNA extracts were stored at -20°C when not in use.

Note: The samples were grinded to a fine powder either using a mortar and pestle or TissueLyser. The lyophilized samples were grinded successfully with both methods, but TissueLyser was not a good choice for fresh samples. Instead of getting a fine powder, the TissueLyser was turning fresh material into squashy product.

2.1.3 Primers selection

DNA sequences from several sources were used to design oligonucleotide primers for amplifying ALS gene fragments (White et al., 2003, Kolkman et al., 2004). Two primers were designed using the software Primer 3. The primers Hel ForA (CAATGGAGATCCACCAAGCT) and Hel RevA (AACGCAAGCAACAAATCACT) used for amplification approximately 700bp fragments.

Note: In the literature, there are plenty of primers, which can be used to detect mutations responsible for resistance/tolerance of different sunflower forms to herbicides. Based on their analysis and comparison, and analysis of sequences of DNA fragments from sunflower stored at the GenBank new primers were designed.

2.1.4 Amplification of specific region of the ALS gene

Final PCR reaction condition were: 19 μ l of mastermix (10 units Biomix, 7 units DEPC water, 1 unit forward primer and 1 unit reverse primer) and 1 μ l of DNA sample. Cycling conditions were: 2 min incubation at 94°C; 35 cycles of 30 sec denaturation at 94°C, 20 sec annealing at 53°C and 45 sec extension at 72°C; and 5 min final extension at 72°C. PCR products were electrophoresed on 2% low-melt agarose gel containing ethidium bromide.

Note: Amplification of DNA fragments was successful while using Biomix, purchased from one manufacturer. Switching to other manufacturer of Biomix failed to generate any PCR products. Initially, we didn't realize what was causing the problem and spent significant time and materials, checking other components of PCR reaction and optimizing the assay itself, but without success. Finally, when we changed Biomix again, and chose the one that had been initially used, the amplification became successful again.

2.1.5 Sequencing

PCR products purification was done before sequencing using the Spin Column PCR Purification Kit following the manufacturer's protocol (http://www.nbsbio.co.uk/downloads/DNA_Cleanup_Handbook.pdf). Purified products were sent together with the corresponding primer (Hel ForA) to Sorce Bioscience (Osford, UK) for sequencing. Analysis of obtained sequences were done based

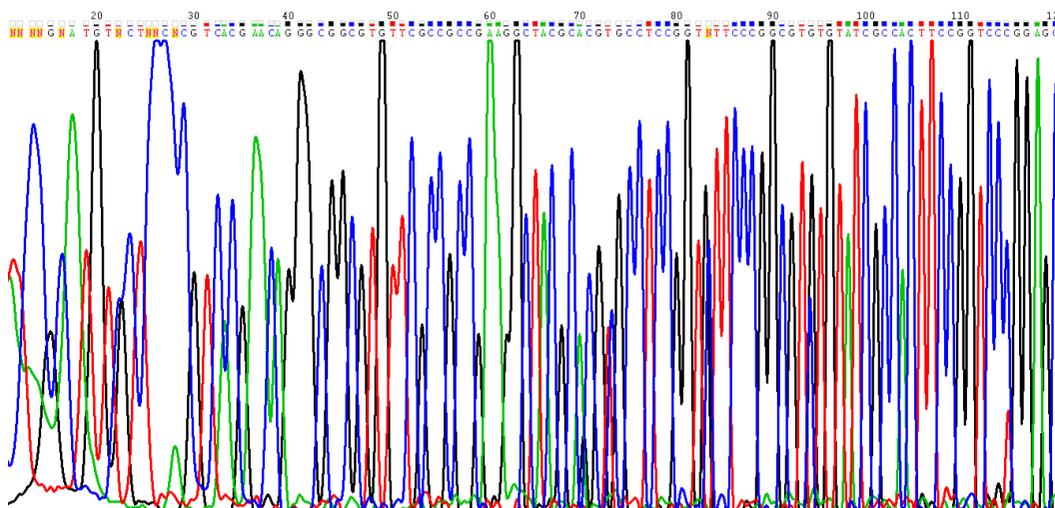


Figure 1: Chromatogram from repeated sequencing of the region of the ALS gene in weedy sunflower DNA.

on comparison with sequences of the amplified region of ALS gene located in GenBank using a multiple sequence alignment program Clustal Omega.

Note: Several sequences obtained from Source Bioscience were not readable. Therefore, it was necessary to repeat sequencing (Figure 1).

2.2 Multiplex PCR-based analysis of microsatellites in three weedy sunflower populations

Multiplex PCR-based analysis of microsatellites is suitable for studies population variability of weeds. Namely, variation in satellite DNA sequences (different size repeated DNA sequences) can be used to determine genetic differences between organisms or closely related individuals (e.g. weedy sunflower which is result of hybridization between different sunflower forms including crop plant, off-type plants, wild plants, volunteer plants and weedy forms). Satellite loci can be defined by the length of the core repeat, number of repeats or the overall repeat length. Microsatellites are DNA repeats with 2–6 nucleotides in length and they are also called simple sequence repeats (SSRs) or short tandem repeats (STRs).

2.2.1 Plant material

Seeds of three different population of weedy sunflower were collected and sown in the greenhouse. Seedlings were transplanted into larger pots. Fresh leaf material from 10 randomly selected plants from each population was collected for DNA extraction. Some leaf samples (taken from a single plant) were used for DNA extraction immediately after sampling, while some them stored in a freezer (−20°C) until analysis. Before analysis samples were lyophilized after storage at the −80°C during 24h.

2.2.2 DNA extraction

DNA was isolated from about 100 mg of fresh plant leaves according to the QiagenDneasy® Plant Mini Kit following the manufacturer's protocol (<https://www.qiagen.com/dz/shop/sample-technologies/dna/dna-preparation/dneasy-plant-mini-kit/>). The quality and concentration of extracted DNA were determined spectrophotometrically using a Nanodrop® 1000. DNA extracts were stored at −20°C when not in use.

Note: Yield of DNA from lyophilized samples was low and curve on Nanodrop® 1000 was unacceptable. Possible reason was high concentration of proteins. To avoid that, we tried to add polyvinylpyrrolidone (PVP), but the problem persisted. Extraction was repeated using fresh leaf samples and satisfactory yield of DNA was obtained.

2.2.3 Primers selection

Seven microsatellite loci were selected from Garayalde et al. (2011) and Muller et al. (2010). The SSR fluorescently labelled markers were sorted by allele-length range (Table 1).

Marker Name	Forward primer sequence	Reverse primer sequence	Allele size range
ORS297	FAM-GTGTCTGCACGAAGCTGTGGT	TGCAAAGCTCACACTAACCTG	214–237
ORS309	FAM-CATTTGGATGGAGCCACTTT	GATGAAGATGGGGAATTTGTG	116–130
ORS337	FAM-TTGGTTCATTCATCCTTGGTG	GGGTGGTGGTTAATTCGTC	165–197
ORS342	NED-TGTTTCATCAGGTTGTCTCCA	CACCAGCATAGCCATTCAAAA	305–361
ORS371	HEX-GGTGCCTTCTCTCCTTGTG	CACACCACCAAACATCAACC	234–264
ORS432	HEX-TGGACCAGTCGTAATCTTTGC	AAACGCATGCAAATGAGGAT	155–167
ORS656	NED-TCGTGGTAAGGGAAGACAACA	ACGGACGTAGAGTGGTGGAG	181–254

Table 1: Microsatellite loci, fluorescent dye, sequence, allele size range for 7 SSR markers.

The amplification reaction were examined for each primer separately consisted of 0.1 μ l of reverse primer and 0.1 μ l of forward primer, fluorescently labelled with NED, HEX or FAM, 5 μ l MMx2 (Taq), 3.8 μ l Rnase-free water and 1 μ l template DNA in a total volume of 10 μ l. Also, the actual amplification reaction for 7 primer together consisted of each unlabelled reserve primer (7 \times 0.1 μ l) and each of forward primer (7 \times 0.1 μ l), 5 μ l MM \times 2 (Taq), 2.6 μ l H₂O and 1 μ l template DNA a total volume of 10 μ l.

2.2.4 PCR analysis

PCR was done following the manufacturer's protocol Type-it®Microsatellite PCR Handbook (<https://www.qiagen.com/dz/resources/search-resources>).

Thermal Cycler (Applied Biosystems Verite 95 Well) was programmed for initial denaturation step of 94°C for 5 min, followed by 6 touchdown cycles of 94°C for 30 s, touchdown annealing temperature (Tx) for 90 s (Tx is initially 63°C and decreases of 1°C per cycle for the six first cycles, until it reaches 57°C) and 72°C for 60 s. PCR products were subsequently amplified for 29 cycles at 94°C for 30 s, touchdown annealing temperature 57°C for 90 s and 72°C for 60 s with a final extension at 63°C for 30 min. DNA Fragment Analysis by the Capillary Electrophoresis system is done in Source Bioscience (Nottingham, UK).

Data analysis

GENEMAPPER (Applied biosystem) and PEAK SCANNER software were used for analyses of the DNA fragments and to score the genotypes (Figure 2).

Sample File	Analysis Method	Panel	Size Standard	Run Name	Run Date & Time	REF	SGI	SFNF	M/F	SNF	SO
01-A01_A1_A01.fsa	Sunflower Plate 1	None	CSLZ500	Sunflower Plate 1	2015-07-22 13:49:05						
02-B01_B1_B01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
03-C01_C1_C01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
04-D01_D1_D01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
05-E01_E1_E01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
06-F01_F1_F01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
07-G01_G1_G01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
08-H01_H1_H01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
09-I01_I1_I01.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
10-E02_E2_E02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
11-C02_C2_C02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
12-D02_D2_D02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
13-E02_E2_E02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
14-F02_F2_F02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
15-G02_G2_G02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
16-H02_H2_H02.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
17-A03_A3_A03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
18-B03_B3_B03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
19-C03_C3_C03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
20-D03_D3_D03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
21-E03_E3_E03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
22-F03_F3_F03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
23-G03_G3_G03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
24-H03_H3_H03.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
25-A04_A4_A04.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
26-B04_B4_B04.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
27-C04_C4_C04.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
28-D04_D4_D04.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
29-E04_E4_E04.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						
30-F04_F4_F04.fsa	Sunflower Plate 1	None	None	Sunflower Plate 1	2015-07-22 13:49:05						

Figure 2: Analyses of the DNA fragments using GENEMAPPER.

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